

DATASET BRIEF

Proteomic analysis of the phytopathogenic fungus *Botrytis cinerea* during cellulose degradation

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The ascomycete *Botrytis cinerea* is a phytopathogenic fungus infecting and causing significant yield losses in a number of crops. Moreover, in the last few years, *B. cinerea* has been adopted as an important model system in molecular phytopathology. In spite of these contributions, the molecular basis of the infection cycle remains unclear. Proteomic approaches have revealed significant information about the infective cycle of several pathogens, including *B. cinerea*. The main aim of this study is to make available a proteomic database containing a significant number of identified proteins from *B. cinerea*. In brief, three independent *B. cinerea* cultures supplemented with carboxymethylcellulose were used, and the extracted proteins were independently separated by 2-D PAGE to obtain the proteome map from *B. cinerea*. Two hundred and sixty-seven spots were selected for MALDI TOF/TOF MS analysis, resulting in 303 positive identifications, mostly representing unannotated proteins. Identified proteins were then classified into categories using the PANTHER classification system (www.pantherdb.org), showing the relevance of protein metabolism and modification process and oxidoreductase activity. Since cellulose is one of the major components of the plant cell wall, many of the identified proteins may have a crucial role in the pathogenicity process. In brief, this proteomic map of *B. cinerea* will be a useful basis for exploring the proteins involved in the infection cycle, which will in turn provide new targets for crop diagnosis and focused fungicide design.

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In the last few years, *Botrytis cinerea* has been adopted as an important model system in molecular phytopathology. Several approaches have been applied to this fungus to unravel its mechanism of infection [1]. These studies have revealed a wide variety and complexity of infection strategies, such as the use of active oxygen species (AOS), toxin

production and the production of cell wall degrading enzymes, among others. In spite of these findings, neither the molecular basis of the synthesis and production of different virulence factors nor the mechanisms of phytopathogenicity used by this fungus are yet well understood [2, 3]. These “black boxes” are being studied by more than a dozen teams [4], improving transformation protocols, vectors, mutants, *etc.* but few articles have been published about the proteins produced by *B. cinerea* [3, 5, 6]. This study is intended to establish a proteomic map of *B. cinerea* during cellulose degradation. The ability of this fungus to secrete a large number of cell wall degrading enzymes has been well studied [7]. Though genes for some cell wall degrading enzymes involved in the degradation of plant cell wall have been characterized, their role in pathogenesis has not been

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Abbreviations: AOS, active oxygen species; CMC, carboxymethylcellulose; TCA, trichloroacetic acid

deeply studied in *B. cinerea*. For this reason we have focused our study on these proteins.

Recently, a large amount of genomic information has become available, including the DNA sequences of *B. cinerea* (strains B05.10 and T4) and its neighbor species *Sclerotinia sclerotinium* [8]. These resources have stimulated the development of high-throughput functional analyses such as transcriptomics and proteomics. Studies in *Saccharomyces cerevisiae* have clearly demonstrated the complementary nature of these approaches by revealing significant differences between the amounts of transcripts and the levels of protein expression [9]. Although protein function cannot always be predicted from gene sequences, especially in organism-specific pathways, ongoing DNA sequencing projects provide a tremendous resource for proteomics, facilitating the generation of protein catalogs correlated to samples or phenotypes for future data mining [10]. The intensive description of the protein 2-DE map of *B. cinerea* may elucidate the “black boxes” of its biology, infection strategies and life cycle. Moreover, the identification of proteins expressed will provide a large number of target candidates to design new molecules for disease control. Despite the increasing number of publications providing molecular information about the pathogenicity mechanisms of *B. cinerea*, at present, there is no fungicide on the market that was developed on the basis of targeted molecular research [4]. The characterization of the *B. cinerea* proteome may result in new targets for antifungal drug screening. Moreover, the definition of a *B. cinerea* 2-DE map provides a basis for comparing proteome analyses between labs.

B. cinerea strain 2100 was obtained from the Spanish Type Culture Collection. Conidial stock suspensions were prepared and conserved as reported previously [11]. The whole fungal proteome of *B. cinerea* was prepared from fungal mycelia from 5-day-old liquid cultures. Three 250 mL flasks each containing 100 mL of minimal salts medium (50 mM NH_4Cl , 7.3 mM KH_2PO_4 , 4.2 mM MgSO_4 , 6.7 mM KCl and 0.07 mM FeSO_4) [12] supplemented with 1% carboxymethylcellulose (CMC) (Panreac, Barcelona, Spain) were inoculated with three 0.5 cm agar plugs taken from the outer edge of 3-day-old colonies of fungal mycelia on potato dextrose agar (Becton Dickinson, Sparks, MD, USA). To check the existence of a specific fungal response to cellulose, a control experiment using glucose as a sole carbon source was performed in parallel. The cultures were grown at 22°C with agitation (180 rpm) for 5 days; mycelia were harvested by filtration and washed with sterile water. A total protein fraction was isolated using a phenol-based procedure outlined previously [13] with some modifications. In short, three independent replicates of mycelia (2 g apiece) were ground to a fine powder under liquid nitrogen. The powder was transferred to 15 mL tubes and resuspended in 10 mL of cold acetone. After vortexing for 30 s, the suspension was centrifuged ($10\,000 \times g$, 5 min, 4°C), and the resulting pellet was washed once again with acetone and resuspended in 10 mL 20% w/v trichloroacetic acid (TCA) in acetone. After centrifugation, the pellets were sequentially washed twice with 20% w/v TCA in acetone, once with 20% w/v TCA and finally twice with 80% v/v acetone. This pellet was air-dried and the dry powder was resuspended in 5 mL “dense SDS buffer” (30% w/v sucrose, 2% w/v SDS, 0.1 M Tris-HCl, pH 8.0, 5% v/v 2-mercaptoethanol). Then, 5 mL Tris-

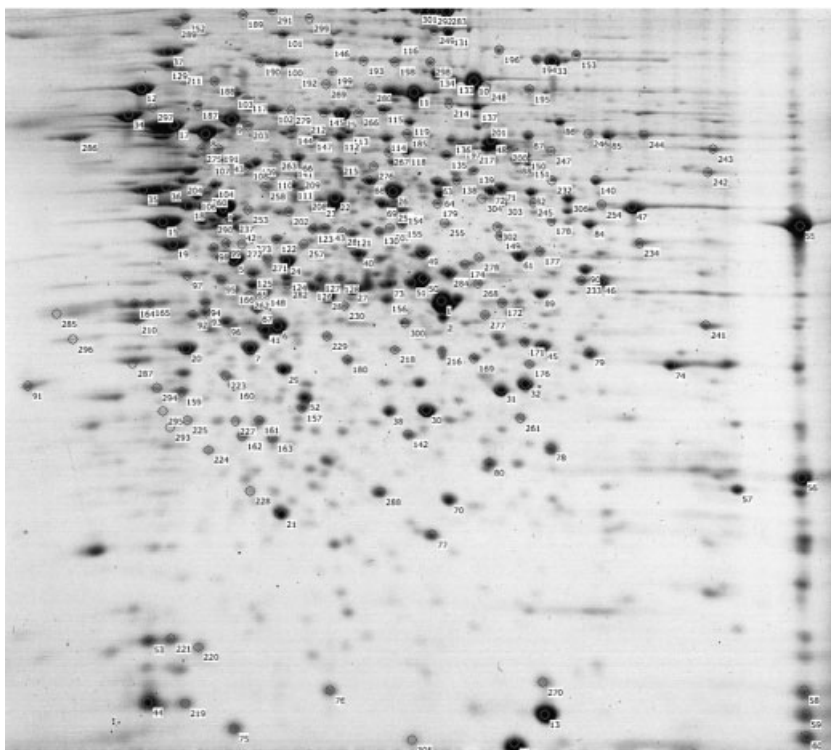


Figure 1. 2-DE CBB-stained gels from mycelial extracts of *B. cinerea* 2100 growing in CMC. Proteins were separated on 17 cm, pH 3–10 non-linear gradient IPG gels (IEF) and 14% polyacrylamide gels (SDS-PAGE).

Table 1. List of identified proteins from *Botrytis cinerea* growing in CMC

| Spot no. | Acc. number | pI | MW | Cov (%) | Mascot MS | Mascot MS/MS | MS/MS peptides | Protein |
|----------|-------------|-----|------|---------|-----------|--------------|----------------|--|
| 1 | BC1G_11968 | 5.9 | 36.7 | 75.8 | 280 | 241 | 2 | <i>B. cinerea</i> glyceraldehyde 3-phosphate dehydrogenase (339 nt) |
| 2 | BC1G_11968 | 5.9 | 36.7 | 80.5 | 320 | 69 | 2 | <i>B. cinerea</i> glyceraldehyde 3-phosphate dehydrogenase (339 nt) |
| 4 | BC1G_00350 | 5.1 | 47.3 | 66.1 | 177 | 318 | 2 | <i>B. cinerea</i> enolase (439 nt) |
| 5 | BC1G_08198 | 5.4 | 41.7 | 65.2 | 246 | 166 | 2 | <i>B. cinerea</i> actin (376 nt) |
| 6 | BC1G_04836 | 5.3 | 39.3 | 75.6 | 233 | 178 | 1 | <i>B. cinerea</i> fructose-bisphosphate aldolase (361 nt) |
| 7 | BC1G_16357 | 5.1 | 23.8 | 89.4 | 177 | 301 | 2 | <i>B. cinerea</i> short chain dehydrogenase (218 nt) |
| 8 | BC1G_10846 | 5.1 | 46 | 64 | 226 | 172 | 2 | <i>B. cinerea</i> hypothetical protein similar to hsp70 (425 nt) |
| 9 | BC1G_11661 | 5.7 | 73.5 | 52.4 | 314 | 198 | 2 | <i>B. cinerea</i> hypothetical protein similar to 70 kDa HSP (680 nt) |
| 10 | BC1G_12307 | 6 | 86.3 | 71.1 | 392 | 122 | 2 | <i>B. cinerea</i> hypothetical protein similar to cobalamin-independent methionine synthase (768 nt) |
| 11 | BC1G_16294 | 4.9 | 34.6 | 54.9 | 130 | 123 | 2 | <i>B. cinerea</i> hypothetical protein (319 nt) |
| 12 | BC1G_00350 | 5.1 | 47.3 | 37.8 | 60 | 206 | 2 | <i>B. cinerea</i> enolase (439 nt) |
| 12 | BC1G_07315 | 4.7 | 79.6 | 58.3 | 289 | 137 | 2 | <i>B. cinerea</i> hypothetical protein similar to hsp84 (702 nt) |
| 13 | BC1G_12729 | 6.6 | 18.2 | 94.6 | 215 | 181 | 2 | <i>B. cinerea</i> woronin body major protein (167 nt) |
| 14 | BC1G_01740 | 6.4 | 19.7 | 72.5 | 108 | 160 | 2 | <i>B. cinerea</i> peptidyl-prolyl cis-trans isomerase (182 nt) |
| 15 | BC1G_11962 | 4.6 | 21.1 | 79.8 | 71 | 18 | 1 | <i>B. cinerea</i> ATP synthase beta chain (193 nt) |
| 17 | BC1G_04390 | 4.8 | 73.2 | 1.8 | x | 100 | 1 | <i>B. cinerea</i> dnaK-type molecular chaperone BiP (671 nt) |
| 17 | BC1G_06164 | 4.8 | 59.9 | 68.8 | 254 | 125 | 2 | <i>B. cinerea</i> heat shock 70 kDa protein 2 (551 nt) |
| 18 | BC1G_12086 | 5 | 54.4 | 36.6 | 62 | x | x | <i>B. cinerea</i> hexokinase (492 nt) |
| 19 | BC1G_00466 | 5 | 45 | 57.9 | 180 | 216 | 3 | <i>B. cinerea</i> eukaryotic initiation factor 4A (399 nt) |
| 20 | BC1G_00555 | 4.2 | 11.2 | 84.2 | 81 | 127 | 2 | <i>B. cinerea</i> transaldolase (101 nt) |
| 21 | BC1G_08882 | 5.4 | 27.1 | 89.9 | 230 | 296 | 2 | <i>B. cinerea</i> hypothetical protein (248 nt) |
| 22 | BC1G_06362 | 5.7 | 53.7 | 73 | 348 | 199 | 2 | <i>B. cinerea</i> hypothetical protein (497 nt) |
| 23 | BC1G_05991 | 5.4 | 51.4 | 67 | 327 | 201 | 2 | <i>B. cinerea</i> hypothetical protein similar to ATP-citrat-lyase (476 nt) |
| 24 | BC1G_03241 | 5.5 | 48.9 | 66 | 285 | 177 | 2 | <i>B. cinerea</i> -conserved hypothetical protein (450 nt) |
| 25 | BC1G_08475 | 5.4 | 67 | 64.4 | 339 | 205 | 2 | <i>B. cinerea</i> -conserved hypothetical protein (613 nt) |
| 26 | BC1G_02744 | 5.9 | 54.8 | 65.7 | 261 | 111 | 2 | <i>B. cinerea</i> 6-phosphogluconate dehydrogenase (495 nt) |
| 27 | BC1G_05297 | 6.2 | 46.5 | 44.9 | 125 | 113 | 2 | <i>B. cinerea</i> hypothetical protein (408 nt) |
| 28 | BC1G_04443 | 9.3 | 44.3 | 67.9 | 309 | 114 | 2 | <i>B. cinerea</i> ketol-acid reductoisomerase. Mitochondrial precursor (399 nt) |
| 29 | BC1G_11823 | 6.7 | 26.6 | 64.4 | 140 | 79 | 2 | <i>B. cinerea</i> inorganic pyrophosphatase (236 nt) |
| 30 | BC1G_07409 | 8.9 | 35.7 | 86.3 | 286 | 142 | 2 | <i>B. cinerea</i> -conserved hypothetical protein (342 nt) |
| 31 | BC1G_10724 | 6.2 | 34.5 | 77.5 | 268 | 207 | 2 | <i>B. cinerea</i> -conserved hypothetical protein (333 nt) |
| 32 | BC1G_10054 | 6.6 | 35 | 93.7 | 356 | 249 | 2 | <i>B. cinerea</i> guanine nucleotide-binding protein beta subunit (317 nt) |
| 33 | BC1G_09731 | 5.9 | 85.9 | 60.1 | 316 | 174 | 2 | <i>B. cinerea</i> hypothetical protein similar to elongation factor 2 (775 nt) |
| 34 | BC1G_04390 | 4.8 | 73.2 | 59.5 | 323 | 261 | 2 | <i>B. cinerea</i> dnaK-type molecular chaperone BiP (671 nt) |
| 35 | BC1G_05299 | 4.8 | 54.2 | 48.1 | 214 | 173 | 2 | <i>B. cinerea</i> hypothetical protein (489 nt) |
| 36 | BC1G_05299 | 4.8 | 54.2 | 47.4 | 166 | 111 | 2 | <i>B. cinerea</i> hypothetical protein (489 nt) |
| 36 | BC1G_05600 | 4.8 | 50.1 | 54.8 | 165 | 82 | 1 | <i>B. cinerea</i> tubulin alpha chain (451 nt) |
| 37 | BC1G_07510 | 4.9 | 90.1 | 49.5 | 260 | 194 | 2 | <i>B. cinerea</i> hypothetical protein similar to valosin-containing protein (824 nt) |
| 38 | BC1G_07233 | 8.8 | 32.2 | 63.4 | 171 | 143 | 2 | <i>B. cinerea</i> hypothetical protein (298 nt) |
| 40 | BC1G_12890 | 6.3 | 44.7 | 68.5 | 264 | 232 | 2 | <i>B. cinerea</i> phosphoglycerate kinase (419 nt) |
| 41 | BC1G_15343 | 5.5 | 37.6 | 78.7 | 322 | 154 | 2 | <i>B. cinerea</i> hypothetical protein (347 nt) |
| 42 | BC1G_01334 | 5.9 | 52.9 | 75.1 | 285 | 181 | 2 | <i>B. cinerea</i> mitochondrial processing peptidase beta subunit. Mitochondrial precursor (481 nt) |
| 43 | BC1G_04759 | 5.6 | 55 | 58.5 | 252 | 239 | 2 | <i>B. cinerea</i> -conserved hypothetical protein (511 nt) |
| 44 | BC1G_05503 | 4.9 | 14.6 | 95.2 | 185 | 99 | 1 | <i>B. cinerea</i> hypothetical protein (126 nt) |
| 45 | BC1G_06394 | 6.8 | 37.5 | 70.5 | 183 | 108 | 1 | <i>B. cinerea</i> hypothetical protein (352 nt) |
| 46 | BC1G_00206 | 9.4 | 41.4 | 64.9 | 234 | 52 | 2 | <i>B. cinerea</i> isocitrate dehydrogenase subunit 1. Mitochondrial precursor (379 nt) |
| 46 | BC1G_15577 | 9.2 | 34.7 | 39.8 | 90 | x | x | <i>B. cinerea</i> hypothetical protein (329 nt) |
| 47 | BC1G_07780 | 9.6 | 60.8 | 7.1 | x | 263 | 3 | <i>B. cinerea</i> hypothetical protein similar to mitochondrial ATP synthase alpha subunit (562 nt) |

Table 1. Continued

| Spot no. | Acc. number | pI | MW | Cov (%) | Mascot MS | Mascot MS/MS | MS/MS peptides | Protein |
|----------|-------------|-----|------|---------|-----------|--------------|----------------|--|
| 48 | BC1G_05305 | 6.9 | 56.7 | 54.1 | 181 | 168 | 2 | <i>B. cinerea</i> hypothetical protein (518 nt) |
| 49 | BC1G_04602 | 5.9 | 43.2 | 74 | 300 | 153 | 2 | <i>B. cinerea</i> S-adenosylmethionine synthetase (396 nt) |
| 50 | BC1G_12319 | 8.8 | 27 | 77.2 | 252 | 177 | 2 | <i>B. cinerea</i> NAD-dependent formate dehydrogenase (246 nt) |
| 51 | BC1G_08723 | 6 | 41.8 | 68.2 | 192 | 102 | 2 | <i>B. cinerea</i> hypothetical protein (377 nt) |
| 51 | BC1G_09640 | 5.9 | 38.9 | 66.3 | 175 | 135 | 2 | <i>B. cinerea</i> hypothetical protein (365 nt) |
| 51 | BC1G_12319 | 8.8 | 27 | 67.1 | 71 | 123 | 1 | <i>B. cinerea</i> NAD-dependent formate dehydrogenase (246 nt) |
| 52 | BC1G_06836 | 5.4 | 55.3 | 28.7 | 85 | 94 | 2 | <i>B. cinerea</i> hypothetical protein (522 nt) |
| 53 | BC1G_09782 | 4.9 | 17.9 | 18.7 | 26 | 126 | 2 | <i>B. cinerea</i> translation initiation factor eIF-5A (166 nt) |
| 55 | BC1G_09492 | 9.8 | 50.1 | 46.9 | 136 | 110 | 2 | <i>B. cinerea</i> elongation factor 1-alpha (461 nt) |
| 56 | BC1G_02099 | 10 | 34 | 61.5 | 127 | 272 | 3 | <i>B. cinerea</i> ADP/ATP carrier protein (314 nt) |
| 56 | BC1G_15613 | 11 | 29.4 | 77.2 | 200 | 56 | 1 | <i>B. cinerea</i> 40S ribosomal protein S4 (263 nt) |
| 57 | BC1G_04187 | 9.4 | 30.1 | 89.8 | 291 | 111 | 2 | <i>B. cinerea</i> outer mitochondrial membrane protein porin (284 nt) |
| 58 | BC1G_03437 | 10 | 17 | 64.2 | 99 | 77 | 2 | <i>B. cinerea</i> 40S ribosomal protein S17 (148 nt) |
| 58 | BC1G_12468 | 11 | 20.8 | 83.4 | 139 | 85 | 2 | <i>B. cinerea</i> 60S ribosomal protein L18 (175 nt) |
| 58 | BC1G_15154 | 11 | 18.4 | 50.9 | 33 | 43 | 1 | <i>B. cinerea</i> 60S ribosomal protein L21 (161 nt) |
| 59 | BC1G_00988 | 12 | 18.5 | 66.7 | 134 | 94 | 2 | <i>B. cinerea</i> 40S ribosomal protein S11 (162 nt) |
| 59 | BC1G_08345 | 11 | 20.1 | 46 | 137 | 104 | 2 | <i>B. cinerea</i> 60S ribosomal protein L11 (176 nt) |
| 60 | BC1G_03667 | 9.9 | 17.9 | 36.7 | 33 | 96 | 1 | <i>B. cinerea</i> 60S ribosomal protein L12 (166 nt) |
| 60 | BC1G_15344 | 10 | 19 | 62.4 | 102 | 44 | 1 | <i>B. cinerea</i> 40S ribosomal protein S10 (170 nt) |
| 61 | BC1G_02443 | 8.7 | 33.2 | 56 | 130 | 109 | 1 | <i>B. cinerea</i> hypothetical protein (302 nt) |
| 63 | BC1G_08348 | 6.6 | 58.5 | 47 | 139 | 188 | 2 | <i>B. cinerea</i> glucose-6-phosphate 1-dehydrogenase (508 nt) |
| 63 | BC1G_11051 | 6.1 | 41.7 | 53.9 | 101 | 124 | 2 | <i>B. cinerea</i> hypothetical protein (401 nt) |
| 64 | BC1G_15108 | 5.6 | 38.2 | 53.1 | 151 | 129 | 2 | <i>B. cinerea</i> hypothetical protein (343 nt) |
| 65 | BC1G_00185 | 5.1 | 39.7 | 41.6 | 135 | 181 | 2 | <i>B. cinerea</i> predicted protein (368 nt) |
| 66 | BC1G_11347 | 5.6 | 62.8 | 67.5 | 294 | 289 | 2 | <i>B. cinerea</i> hypothetical protein (573 nt) |
| 67 | BC1G_08819 | 5.4 | 37.4 | 85.2 | 195 | 23 | 1 | <i>B. cinerea</i> fructose-1,6-bisphosphatase (344 nt) |
| 67 | BC1G_14880 | 5.4 | 39.4 | 69.6 | 247 | 76 | 2 | <i>B. cinerea</i> hypothetical protein (369 nt) |
| 68 | BC1G_06102 | 5.5 | 30.7 | 46 | 74 | 104 | 2 | <i>B. cinerea</i> hypothetical protein (272 nt) |
| 68 | BC1G_06103 | 5.6 | 53.6 | 43 | 127 | x | x | <i>B. cinerea</i> hypothetical protein (472 nt) |
| 68 | BC1G_08348 | 6.6 | 58.5 | 51.8 | 165 | 176 | 2 | <i>B. cinerea</i> glucose-6-phosphate 1-dehydrogenase (508 nt) |
| 69 | BC1G_11376 | 6.7 | 54.5 | 45 | 213 | 195 | 2 | <i>B. cinerea</i> dihydrolipoyl dehydrogenase. Mitochondrial precursor (511 nt) |
| 70 | BC1G_00059 | 6.1 | 25.7 | 77.4 | 330 | 117 | 2 | <i>B. cinerea</i> -conserved hypothetical protein (230 nt) |
| 71 | BC1G_01968 | 6.3 | 57.6 | 80 | 366 | 199 | 2 | <i>B. cinerea</i> hypothetical protein (510 nt) |
| 72 | BC1G_10557 | 6.6 | 58.7 | 78.7 | 335 | 158 | 2 | <i>B. cinerea</i> hypothetical protein similar to UDP-glucose pyrophosphorylase (526 nt) |
| 73 | BC1G_10112 | 5.7 | 41.5 | 67.5 | 148 | 40 | 2 | <i>B. cinerea</i> cyanide hydratase (372 nt) |
| 74 | BC1G_03920 | 9.2 | 38.2 | 76.1 | 309 | 166 | 2 | <i>B. cinerea</i> hypothetical protein (339 nt) |
| 75 | BC1G_05133 | 5.2 | 16.4 | 39.5 | 71 | 192 | 2 | <i>B. cinerea</i> hypothetical protein (157 nt) |
| 76 | BC1G_14488 | 6.4 | 21.9 | 50 | 156 | 177 | 2 | <i>B. cinerea</i> hypothetical protein (192 nt) |
| 77 | BC1G_00383 | 6.3 | 24.6 | 71.7 | 150 | 111 | 2 | <i>B. cinerea</i> GTP-binding nuclear protein ran-1 (219 nt) |
| 78 | BC1G_04230 | 7.7 | 30.9 | 72.8 | 261 | 141 | 2 | <i>B. cinerea</i> hypothetical protein (290 nt) |
| 79 | BC1G_14078 | 6.9 | 35.8 | 76.1 | 248 | 185 | 2 | <i>B. cinerea</i> hypothetical protein (331 nt) |
| 80 | BC1G_04783 | 6.5 | 24.5 | 47.7 | 111 | 194 | 2 | <i>B. cinerea</i> hypothetical protein (235 nt) |
| 82 | BC1G_10557 | 6.6 | 58.7 | 75.1 | 269 | 126 | 2 | <i>B. cinerea</i> hypothetical protein similar to UDP-glucose pyrophosphorylase (526 nt) |
| 82 | BC1G_12462 | 6 | 28.8 | 68.1 | 94 | 277 | 3 | <i>B. cinerea</i> hypothetical protein (254 nt) |
| 84 | BC1G_06851 | 7.8 | 52.8 | 69 | 299 | 205 | 2 | <i>B. cinerea</i> hypothetical protein similar to serine hydroxymethyltransferase (478 nt) |
| 85 | BC1G_05989 | 9.4 | 57.7 | 63.1 | 253 | 114 | 2 | <i>B. cinerea</i> ATP-citrate synthase subunit 1 (531 nt) |
| 86 | BC1G_02144 | 6.7 | 74.4 | 64.9 | 315 | 214 | 2 | <i>B. cinerea</i> hypothetical protein (667 nt) |
| 87 | BC1G_16089 | 9.6 | 52.9 | 40.8 | 171 | 149 | 2 | <i>B. cinerea</i> fumarate reductase flavoprotein subunit (485 nt) |
| 88 | BC1G_12242 | 6.5 | 64.1 | 58.9 | 275 | 143 | 2 | <i>B. cinerea</i> hypothetical protein (574 nt) |
| 88 | BC1G_13616 | 6.4 | 61 | 45.4 | 139 | 61 | 1 | <i>B. cinerea</i> isocitrate lyase (549 nt) |
| 89 | BC1G_13706 | 6.7 | 49 | 56.8 | 241 | 192 | 2 | <i>B. cinerea</i> hypothetical protein (440 nt) |
| 90 | BC1G_02472 | 9.4 | 48.2 | 60.4 | 186 | 249 | 2 | <i>B. cinerea</i> ubiquinol-cytochrome-c reductase complex core protein 2 (462 nt) |
| 91 | BC1G_03337 | 4.6 | 17.8 | 58.5 | 70 | 243 | 2 | <i>B. cinerea</i> hypothetical protein (159 nt) |

Table 1. Continued

| Spot no. | Acc. number | pI | MW | Cov (%) | Mascot MS | Mascot MS/MS | MS/MS peptides | Protein |
|----------|-------------|-----|------|---------|-----------|--------------|----------------|--|
| 92 | BC1G_06110 | 5.1 | 35.6 | 52.2 | 160 | 214 | 2 | <i>B. cinerea</i> hypothetical protein (312 nt) |
| 93 | BC1G_00266 | 5.5 | 34.9 | 43.7 | 96 | 111 | 2 | <i>B. cinerea</i> hypothetical protein (325 nt) |
| 94 | BC1G_00033 | 7.7 | 45.2 | 42 | 100 | 44 | 1 | <i>B. cinerea</i> hypothetical protein (419 nt) |
| 94 | BC1G_01219 | 5.2 | 42.9 | 55.1 | 115 | 332 | 2 | <i>B. cinerea</i> hypothetical protein (381 nt) |
| 95 | BC1G_03473 | 5.5 | 39.4 | 30.1 | 58 | 240 | 3 | <i>B. cinerea</i> hypothetical protein similar to oxaloacetate acetylhydrolase (366 nt) |
| 95 | BC1G_13829 | 5.3 | 44.1 | 61.3 | 200 | 151 | 2 | <i>B. cinerea</i> actin-related protein 2 (390 nt) |
| 96 | BC1G_06571 | 5.3 | 37.8 | 61.9 | 192 | 96 | 1 | <i>B. cinerea</i> adenosine kinase (349 nt) |
| 97 | BC1G_15906 | 5.8 | 48 | 50.4 | 192 | 87 | 2 | <i>B. cinerea</i> ATP-specific succinyl-CoA synthetase beta subunit (446 nt) |
| 98 | BC1G_01354 | 5.2 | 40.9 | 54.4 | 175 | 203 | 2 | <i>B. cinerea</i> saccharopine dehydrogenase (373 nt) |
| 99 | BC1G_12601 | 5.4 | 42.2 | 50 | 125 | 199 | 2 | <i>B. cinerea</i> hypothetical protein (382 nt) |
| 100 | BC1G_00769 | 5.4 | 86 | 37 | 160 | 163 | 2 | <i>B. cinerea</i> hypothetical protein (791 nt) |
| 100 | BC1G_13753 | 4.9 | 46.6 | 37.1 | 65 | x | x | <i>B. cinerea</i> hypothetical protein (428 nt) |
| 101 | BC1G_05626 | 5.6 | 38.6 | 35.4 | 70 | 187 | 3 | <i>B. cinerea</i> hypothetical protein (339 nt) |
| 101 | BC1G_16318 | 8.9 | 23.3 | 70.4 | 94 | 116 | 2 | <i>B. cinerea</i> hypothetical protein (206 nt) |
| 102 | BC1G_06515 | 5.4 | 74.7 | 50.8 | 177 | 230 | 2 | <i>B. cinerea</i> hypothetical protein (666 nt) |
| 103 | BC1G_02363 | 5.2 | 82 | 24.7 | 103 | 55 | 1 | <i>B. cinerea</i> hypothetical protein (728 nt) |
| 103 | BC1G_03729 | 5.3 | 78.8 | 41.7 | 212 | 74 | 1 | <i>B. cinerea</i> hypothetical protein (714 nt) |
| 104 | BC1G_10065 | 5.2 | 51.8 | 44.4 | 136 | 155 | 2 | <i>B. cinerea</i> hypothetical protein (471 nt) |
| 104 | BC1G_11238 | 5.2 | 58.6 | 35.8 | 99 | x | x | <i>B. cinerea</i> hypothetical protein (542 nt) |
| 104 | BC1G_11314 | 5.4 | 40.7 | 4.9 | x | 178 | 1 | <i>B. cinerea</i> hypothetical protein (371 nt) |
| 106 | BC1G_09341 | 5.5 | 61.2 | 49.1 | 187 | 234 | 2 | <i>B. cinerea</i> hsp60 (583 nt) |
| 106 | BC1G_10581 | 5 | 53.2 | 60.5 | 170 | 85 | 2 | <i>B. cinerea</i> -conserved hypothetical protein (479 nt) |
| 107 | BC1G_04759 | 5.6 | 55 | 21.1 | 35 | 91 | 2 | <i>B. cinerea</i> -conserved hypothetical protein (511 nt) |
| 107 | BC1G_14947 | 5.4 | 59.9 | 57.8 | 263 | 136 | 2 | <i>B. cinerea</i> T-complex protein 1. Theta subunit (550 nt) |
| 108 | BC1G_10814 | 5.4 | 56.7 | 48.2 | 187 | 154 | 2 | <i>B. cinerea</i> hypothetical protein (515 nt) |
| 109 | BC1G_04759 | 5.6 | 55 | 55.8 | 208 | 248 | 2 | <i>B. cinerea</i> -conserved hypothetical protein (511 nt) |
| 110 | BC1G_04232 | 5.4 | 57.7 | 75.9 | 354 | 211 | 2 | <i>B. cinerea</i> vacuolar ATP synthase subunit B (518 nt) |
| 111 | BC1G_09363 | 5.5 | 57.4 | 57.2 | 316 | 164 | 2 | <i>B. cinerea</i> hypothetical protein (514 nt) |
| 112 | BC1G_07795 | 6.4 | 72.8 | 71.3 | 332 | 233 | 2 | <i>B. cinerea</i> -conserved hypothetical protein (663 nt) |
| 113 | BC1G_10523 | 5.6 | 119 | 33.7 | 234 | 153 | 2 | <i>B. cinerea</i> hypothetical protein (1048 nt) |
| 114 | BC1G_08339 | 5.9 | 65.1 | 75.1 | 389 | 237 | 2 | <i>B. cinerea</i> hypothetical protein (595 nt) |
| 115 | gil85083792 | 6 | 81.6 | 25.5 | 81 | 121 | 1 | NADH-ubiquinone oxidoreductase 78 kDa subunit precursor (CI-78KD) [<i>Neurospora crassa</i> OR74A] |
| 116 | BC1G_12439 | 6.4 | 34.5 | 69.7 | 143 | 128 | 2 | <i>B. cinerea</i> alpha-ketoglutarate dehydrogenase E1 component (300 nt) |
| 117 | BC1G_12797 | 5.3 | 66.6 | 40.3 | 170 | 124 | 2 | <i>B. cinerea</i> translation initiation factor eIF3 (601 nt) |
| 118 | BC1G_06097 | 5.7 | 52 | 60.5 | 249 | 221 | 2 | <i>B. cinerea</i> hypothetical protein (471 nt) |
| 119 | BC1G_06223 | 5.9 | 70 | 59.4 | 261 | 150 | 2 | <i>B. cinerea</i> hypothetical protein (608 nt) |
| 121 | BC1G_11707 | 5.7 | 44 | 41.4 | 66 | 136 | 2 | <i>B. cinerea</i> translation elongation factor EF-Tu (399 nt) |
| 121 | BC1G_12973 | 9.2 | 58.5 | 53.7 | 139 | 94 | 2 | <i>B. cinerea</i> fumarate hydratase (540 nt) |
| 122 | BC1G_00939 | 5.3 | 47 | 56.3 | 240 | 67 | 1 | <i>B. cinerea</i> hypothetical protein (416 nt) |
| 123 | BC1G_01188 | 5.8 | 38.3 | 56 | 120 | 151 | 2 | <i>B. cinerea</i> hypothetical protein (359 nt) |
| 124 | BC1G_10921 | 9.6 | 57.4 | 34.1 | 96 | 191 | 2 | <i>B. cinerea</i> hypothetical protein (507 nt) |
| 125 | BC1G_11550 | 5.9 | 11.6 | 97.2 | 144 | 219 | 2 | <i>B. cinerea</i> hypothetical protein (108 nt) |
| 125 | gil85792456 | 5.3 | 43 | 84.6 | 386 | 289 | 3 | Mannitol-1-phosphate dehydrogenase [<i>Botryotinia fuckeliana</i>] |
| 126 | BC1G_05297 | 6.2 | 46.5 | 30.6 | 63 | 163 | 3 | <i>B. cinerea</i> hypothetical protein (408 nt) |
| 126 | BC1G_16075 | 5.7 | 29.3 | 51.1 | 107 | 133 | 2 | <i>B. cinerea</i> phosphatidylinositol/phosphatidylcholine transfer protein (264 nt) |
| 127 | BC1G_13607 | 5.8 | 40.3 | 44.6 | 92 | 90 | 1 | <i>B. cinerea</i> peptidyl-prolyl <i>cis</i> - <i>trans</i> isomerase (372 nt) |
| 127 | BC1G_15180 | 5.6 | 40.2 | 29.3 | 49 | 42 | 1 | <i>B. cinerea</i> L-arabinitol 4-dehydrogenase (375 nt) |
| 128 | BC1G_14194 | 5.7 | 41.1 | 57.7 | 189 | 271 | 2 | <i>B. cinerea</i> hypothetical protein (383 nt) |
| 129 | BC1G_09769 | 4.9 | 79.3 | 39.2 | 144 | 140 | 2 | <i>B. cinerea</i> hypothetical protein (712 nt) |
| 130 | BC1G_02852 | 8.8 | 45.6 | 66.1 | 237 | 231 | 2 | <i>B. cinerea</i> hypothetical protein (410 nt) |
| 131 | BC1G_15638 | 6.2 | 106 | 35.9 | 201 | 101 | 2 | <i>B. cinerea</i> translation elongation factor eEF-3 (947 nt) |
| 133 | BC1G_12307 | 6 | 86.3 | 44.5 | 163 | 122 | 2 | <i>B. cinerea</i> hypothetical protein similar to cobalamin-independent methionine synthase (768 nt) |

Table 1. Continued

| Spot no. | Acc. number | pI | MW | Cov (%) | Mascot MS | Mascot MS/MS | MS/MS peptides | Protein |
|----------|--------------|-----|------|---------|-----------|--------------|----------------|--|
| 134 | BC1G_01508 | 6 | 92.9 | 49.4 | 330 | 193 | 2 | <i>B. cinerea</i> phosphoketolase (822 nt) |
| 135 | BC1G_06374 | 6.5 | 53.9 | 66 | 318 | 187 | 2 | <i>B. cinerea</i> hypothetical protein (477 nt) |
| 136 | BC1G_05305 | 6.9 | 56.7 | 50.4 | 167 | 79 | 2 | <i>B. cinerea</i> hypothetical protein (518 nt) |
| 137 | BC1G_07090 | 5.9 | 72.2 | 39.9 | 202 | 214 | 2 | <i>B. cinerea</i> hypothetical protein (652 nt) |
| 138 | BC1G_04076 | 6.2 | 53.8 | 53.5 | 248 | 113 | 2 | <i>B. cinerea</i> hypothetical protein (499 nt) |
| 139 | BC1G_00483 | 6.4 | 55.7 | 46.8 | 207 | 68 | 2 | <i>B. cinerea</i> hypothetical protein (494 nt) |
| 140 | BC1G_09443 | 7.8 | 60.9 | 64.8 | 273 | 116 | 2 | <i>B. cinerea</i> malate synthase (543 nt) |
| 141 | BC1G_12883 | 5.7 | 61.9 | 47.8 | 223 | 205 | 2 | <i>B. cinerea</i> hypothetical protein (540 nt) |
| 142 | BC1G_02778 | 6.1 | 31.7 | 57.5 | 182 | 209 | 2 | <i>B. cinerea</i> hypothetical protein (285 nt) |
| 143 | BC1G_02852 | 8.8 | 45.6 | 63.4 | 235 | 226 | 2 | <i>B. cinerea</i> hypothetical protein (410 nt) |
| 144 | BC1G_05616 | 5.5 | 73.3 | 50.1 | 240 | 236 | 2 | <i>B. cinerea</i> predicted protein (681 nt) |
| 145 | BC1G_08475 | 5.4 | 67 | 44.5 | 194 | 207 | 2 | <i>B. cinerea</i> -conserved hypothetical protein (613 nt) |
| 146 | BC1G_01538 | 5.5 | 106 | 56 | 374 | 164 | 2 | <i>B. cinerea</i> hypothetical protein (953 nt) |
| 147 | BC1G_14651 | 5.5 | 62.1 | 38 | 192 | 232 | 2 | <i>B. cinerea</i> phosphoenolpyruvate carboxykinase (561 nt) |
| 148 | BC1G_12011 | 5.5 | 40.7 | 41.2 | 115 | 267 | 2 | <i>B. cinerea</i> glutamine synthetase (369 nt) |
| 149 | BC1G_02594 | 6.9 | 45.5 | 5.4 | x | 130 | 2 | <i>B. cinerea</i> hypothetical protein (429 nt) |
| 150 | BC1G_12988 | 6.9 | 112 | 37.4 | 271 | 123 | 1 | <i>B. cinerea</i> hypothetical protein (1045 nt) |
| 151 | BC1G_07479 | 9.1 | 65.7 | 48.4 | 275 | 188 | 2 | <i>B. cinerea</i> -conserved hypothetical protein (589 nt) |
| 153 | BC1G_00557 | 9 | 105 | 42.4 | 189 | 68 | 2 | <i>B. cinerea</i> hypothetical protein (940 nt) |
| 153 | BC1G_06628 | 6.5 | 67.2 | 35 | 102 | 121 | 2 | <i>B. cinerea</i> hypothetical protein similar to septin 2 (588 nt) |
| 154 | BC1G_13490 | 5.9 | 49.1 | 61.4 | 336 | 144 | 2 | <i>B. cinerea</i> NADP-specific glutamate dehydrogenase (451 nt) |
| 155 | BC1G_15049 | 5.9 | 49.4 | 31.8 | 153 | 138 | 2 | <i>B. cinerea</i> saccharopine reductase (449 nt) |
| 156 | BC1G_11968 | 5.9 | 36.7 | 71.7 | 203 | 150 | 2 | <i>B. cinerea</i> glyceraldehyde 3-phosphate dehydrogenase (339 nt) |
| 157 | BC1G_08301 | 9.2 | 40.8 | 14.5 | x | 415 | 3 | <i>B. cinerea</i> ascorbate peroxidase (373 nt) |
| 159 | BC1G_04780 | 4.9 | 34.1 | 63.2 | 223 | 115 | 2 | <i>B. cinerea</i> hypothetical protein (299 nt) |
| 159 | BC1G_13374 | 5 | 32.5 | 32.2 | 71 | 60 | 1 | <i>B. cinerea</i> hypothetical protein (304 nt) |
| 160 | BC1G_08794 | 7.7 | 40.8 | 40.5 | 138 | 133 | 2 | <i>B. cinerea</i> pyruvate dehydrogenase E1 component beta subunit (373 nt) |
| 161 | BC1G_08983 | 5.3 | 35.4 | 29.9 | 67 | 246 | 2 | <i>B. cinerea</i> hypothetical protein similar to spermidine synthase (311 nt) |
| 162 | BC1G_05298 | 5.4 | 29.5 | 66.7 | 225 | 70 | 2 | <i>B. cinerea</i> hypothetical protein (264 nt) |
| 163 | BC1G_05298 | 5.4 | 29.5 | 66.7 | 206 | 81 | 2 | <i>B. cinerea</i> hypothetical protein (264 nt) |
| 164 | BC1G_01804 | 4.6 | 31.8 | 63.6 | 189 | 169 | 3 | <i>B. cinerea</i> 40S ribosomal protein S0 (297 nt) |
| 165 | BC1G_01804 | 4.6 | 31.8 | 62.3 | 188 | 181 | 3 | <i>B. cinerea</i> 40S ribosomal protein S0 (297 nt) |
| 166 | BC1G_11454 | 5.2 | 36.3 | 69.4 | 293 | 162 | 2 | <i>B. cinerea</i> hypothetical protein (324 nt) |
| 169 | BC1G_11083 | 6.3 | 36.1 | 60.4 | 161 | 130 | 2 | <i>B. cinerea</i> alcohol dehydrogenase (341 nt) |
| 171 | BC1G_04987 | 6.7 | 38.3 | 91.4 | 344 | 184 | 2 | <i>B. cinerea</i> hypothetical protein (338 nt) |
| 172 | BC1G_06124 | 6.1 | 38.9 | 48.6 | 97 | 218 | 2 | <i>B. cinerea</i> hypothetical protein similar to mannose-1-phosphate guanylyltransferase (354 nt) |
| 172 | gil111065380 | 10 | 50.2 | 6.3 | x | 153 | 2 | Hypothetical protein SNOG_06669 [<i>Phaeosphaeria nodorum</i> SN15] |
| 174 | BC1G_12319 | 8.8 | 27 | 53.7 | 84 | 128 | 1 | <i>B. cinerea</i> NAD-dependent formate dehydrogenase (246 nt) |
| 174 | BC1G_16156 | 10 | 28.6 | 68.3 | 171 | 267 | 2 | <i>B. cinerea</i> NADH-ubiquinone oxidoreductase 40 kDa subunit (252 nt) |
| 176 | BC1G_07285 | 9.3 | 41.7 | 11.7 | 33 | 241 | 3 | <i>B. cinerea</i> hypothetical protein (385 nt) |
| 177 | BC1G_02443 | 8.7 | 33.2 | 8.9 | 22 | 63 | 1 | <i>B. cinerea</i> hypothetical protein (302 nt) |
| 178 | BC1G_07492 | 6.8 | 44.8 | 65.7 | 251 | 195 | 2 | <i>B. cinerea</i> hypothetical protein (417 nt) |
| 179 | BC1G_04985 | 6.1 | 54.4 | 43.1 | 133 | 90 | 1 | <i>B. cinerea</i> hypothetical protein (485 nt) |
| 179 | BC1G_07094 | 6.6 | 56.4 | 31.8 | 60 | x | x | <i>B. cinerea</i> hypothetical protein (493 nt) |
| 180 | BC1G_02348 | 6.1 | 30.9 | 72.6 | 247 | 161 | 2 | <i>B. cinerea</i> hypothetical protein (285 nt) |
| 185 | BC1G_01327 | 5.3 | 50.8 | 44.2 | 176 | 244 | 2 | <i>B. cinerea</i> hypothetical protein (455 nt) |
| 187 | BC1G_07142 | 5.1 | 68.4 | 64.2 | 292 | 183 | 2 | <i>B. cinerea</i> hypothetical protein (617 nt) |
| 188 | BC1G_03226 | 5.1 | 77.1 | 47.8 | 261 | 170 | 2 | <i>B. cinerea</i> hypothetical protein similar to NADPH cytochrome P450 oxidoreductase (693 nt) |
| 189 | BC1G_11655 | 5.8 | 115 | 23.8 | 111 | 113 | 2 | <i>B. cinerea</i> hypothetical protein (1050 nt) |
| 190 | BC1G_02682 | 5.6 | 95 | 50.6 | 288 | 112 | 2 | <i>B. cinerea</i> hypothetical protein (842 nt) |
| 191 | BC1G_06804 | 5.3 | 61.8 | 52.8 | 242 | 76 | 2 | <i>B. cinerea</i> hypothetical protein (591 nt) |
| 192 | BC1G_04169 | 5.5 | 81.5 | 41.7 | 174 | 261 | 2 | <i>B. cinerea</i> hypothetical protein (762 nt) |

Table 1. Continued

| Spot no. | Acc. number | pI | MW | Cov (%) | Mascot MS | Mascot MS/MS | MS/MS peptides | Protein |
|----------|-------------|-----|------|---------|-----------|--------------|----------------|---|
| 193 | BC1G_03007 | 6.2 | 40.3 | 65.4 | 182 | 176 | 2 | <i>B. cinerea</i> hypothetical protein (358 nt) |
| 194 | BC1G_09731 | 5.9 | 85.9 | 49.2 | 216 | 205 | 2 | <i>B. cinerea</i> hypothetical protein similar to elongation factor 2 (775 nt) |
| 195 | BC1G_14312 | 5.4 | 58.3 | 69.4 | 228 | 212 | 2 | <i>B. cinerea</i> hypothetical protein (510 nt) |
| 196 | BC1G_04451 | 6.3 | 101 | 42.8 | 307 | 146 | 2 | <i>B. cinerea</i> hypothetical protein (940 nt) |
| 197 | BC1G_09030 | 6.3 | 58.2 | 54.7 | 248 | 172 | 2 | <i>B. cinerea</i> hypothetical protein (548 nt) |
| 198 | BC1G_03007 | 6.2 | 40.3 | 80.2 | 267 | 161 | 2 | <i>B. cinerea</i> hypothetical protein (358 nt) |
| 199 | BC1G_05496 | 5.7 | 89.4 | 39.9 | 253 | 91 | 2 | <i>B. cinerea</i> hypothetical protein (797 nt) |
| 200 | BC1G_14217 | 6.2 | 58.9 | 62.1 | 281 | 222 | 2 | <i>B. cinerea</i> acetyl-CoA hydrolase (531 nt) |
| 201 | BC1G_04159 | 6.3 | 72.4 | 48.1 | 321 | 118 | 2 | <i>B. cinerea</i> hypothetical protein (643 nt) |
| 202 | BC1G_04300 | 5.6 | 53.6 | 46 | 120 | 210 | 2 | <i>B. cinerea</i> hypothetical protein (487 nt) |
| 202 | BC1G_09350 | 5.5 | 51.5 | 41.6 | 111 | 181 | 3 | <i>B. cinerea</i> -conserved hypothetical protein (471 nt) |
| 203 | BC1G_09514 | 6 | 75.4 | 39.6 | 258 | 131 | 2 | <i>B. cinerea</i> hypothetical protein (670 nt) |
| 204 | BC1G_01381 | 5 | 51.4 | 40.8 | 92 | 90 | 2 | <i>B. cinerea</i> hypothetical protein (468 nt) |
| 204 | BC1G_13687 | 5.4 | 48.4 | 53.4 | 139 | 45 | 1 | <i>B. cinerea</i> dihydrolipoamide acetyltransferase E2 of pyruvate dehydrogenase complex (464 nt) |
| 207 | BC1G_01554 | 5.7 | 47.5 | 34 | 80 | x | x | <i>B. cinerea</i> hypothetical protein (424 nt) |
| 207 | BC1G_03240 | 5.9 | 41.3 | 44.3 | 102 | 193 | 2 | <i>B. cinerea</i> hypothetical protein (368 nt) |
| 207 | BC1G_12069 | 6 | 46.2 | 40.4 | 64 | 98 | 1 | <i>B. cinerea</i> hypothetical protein (416 nt) |
| 208 | BC1G_06362 | 5.7 | 53.7 | 69.2 | 286 | 203 | 2 | <i>B. cinerea</i> hypothetical protein (497 nt) |
| 209 | BC1G_04232 | 5.4 | 57.7 | 68.3 | 361 | 152 | 2 | <i>B. cinerea</i> vacuolar ATP synthase subunit B (518 nt) |
| 210 | BC1G_08895 | 4.8 | 28.8 | 50.4 | 89 | 205 | 2 | <i>B. cinerea</i> hypothetical protein (262 nt) |
| 211 | BC1G_11866 | 4.9 | 85.3 | 39.5 | 145 | 93 | 2 | <i>B. cinerea</i> hypothetical protein (745 nt) |
| 212 | BC1G_06967 | 5.5 | 72.7 | 67 | 403 | 192 | 2 | <i>B. cinerea</i> hypothetical protein similar to fimbrin (651 nt) |
| 214 | BC1G_00347 | 6.3 | 77.7 | 48.4 | 207 | 93 | 2 | <i>B. cinerea</i> hypothetical prot similar to glucosamine-fructose-6-phosphate aminotransferase (702 nt) |
| 214 | BC1G_13832 | 6.4 | 46.3 | 58.5 | 163 | 204 | 3 | <i>B. cinerea</i> hypothetical protein (422 nt) |
| 215 | BC1G_09186 | 6.6 | 65.2 | 49.8 | 265 | 198 | 2 | <i>B. cinerea</i> hypothetical protein (610 nt) |
| 216 | BC1G_04788 | 6.1 | 35.6 | 85.8 | 261 | 271 | 2 | <i>B. cinerea</i> alcohol dehydrogenase (324 nt) |
| 217 | BC1G_07745 | 6.3 | 59.2 | 38.6 | 234 | 112 | 2 | <i>B. cinerea</i> -conserved hypothetical protein (541 nt) |
| 218 | BC1G_06844 | 8.8 | 38 | 49.7 | 143 | 205 | 2 | <i>B. cinerea</i> hypothetical protein (354 nt) |
| 219 | BC1G_07699 | 4.9 | 17.7 | 43.8 | 58 | 182 | 3 | <i>B. cinerea</i> -conserved hypothetical protein (160 nt) |
| 220 | BC1G_05706 | 5.3 | 19.1 | 86.9 | 161 | 16 | 1 | <i>B. cinerea</i> hypothetical protein (198 nt) |
| 221 | BC1G_09782 | 4.9 | 17.9 | 25.9 | 57 | 138 | 2 | <i>B. cinerea</i> translation initiation factor eIF-5A (166 nt) |
| 223 | BC1G_00152 | 6.4 | 25.7 | 68.8 | 125 | 170 | 2 | <i>B. cinerea</i> hypothetical protein (234 nt) |
| 223 | BC1G_07903 | 4.9 | 18.3 | 67.3 | 119 | 257 | 2 | <i>B. cinerea</i> hypothetical protein (165 nt) |
| 223 | BC1G_14077 | 5.1 | 34.3 | 35.3 | 63 | x | x | <i>B. cinerea</i> hypothetical protein (312 nt) |
| 224 | BC1G_15795 | 5.1 | 30.2 | 28.9 | 91 | 92 | 2 | <i>B. cinerea</i> 20S proteasome subunit Y7 (280 nt) |
| 225 | BC1G_02778 | 6.1 | 31.7 | 25.6 | 74 | 128 | 2 | <i>B. cinerea</i> hypothetical protein (285 nt) |
| 225 | BC1G_12159 | 5.1 | 32.7 | 45.4 | 100 | 80 | 1 | <i>B. cinerea</i> hypothetical protein (293 nt) |
| 227 | BC1G_02778 | 6.1 | 31.7 | 54.4 | 112 | 180 | 3 | <i>B. cinerea</i> hypothetical protein (285 nt) |
| 227 | BC1G_09132 | 6.4 | 29.8 | 66.2 | 162 | 96 | 2 | <i>B. cinerea</i> hypothetical protein (272 nt) |
| 228 | BC1G_14431 | 5.5 | 27.6 | 43.5 | 155 | 165 | 2 | <i>B. cinerea</i> glucose 1-dehydrogenase (260 nt) |
| 229 | BC1G_04452 | 5.7 | 37.5 | 79.5 | 315 | 139 | 2 | <i>B. cinerea</i> eukaryotic translation initiation factor 3 (336 nt) |
| 230 | BC1G_10614 | 5.8 | 39.4 | 72.9 | 338 | 143 | 2 | <i>B. cinerea</i> hypothetical protein (351 nt) |
| 232 | BC1G_01297 | 6.6 | 54.5 | 24.4 | 64 | 173 | 3 | <i>B. cinerea</i> hypothetical protein (475 nt) |
| 233 | BC1G_00206 | 9.4 | 41.4 | 32.7 | 70 | x | x | <i>B. cinerea</i> isocitrate dehydrogenase subunit 1. Mitochondrial precursor (379 nt) |
| 233 | BC1G_02655 | 9.4 | 48 | 30.4 | 54 | x | x | <i>B. cinerea</i> acyl-CoA dehydrogenase family protein (438 nt) |
| 233 | BC1G_04778 | 9.2 | 41.1 | 36.5 | 87 | 109 | 2 | <i>B. cinerea</i> GTP-binding protein (370 nt) |
| 234 | BC1G_15535 | 8.6 | 43.9 | 28.4 | 61 | 137 | 3 | <i>B. cinerea</i> hypothetical protein (402 nt) |
| 237 | BC1G_13777 | 5.3 | 52 | 15.3 | 19 | 30 | 1 | <i>B. cinerea</i> secretory pathway Rab GDP dissociation inhibitor (472 nt) |
| 241 | BC1G_15301 | 9.8 | 46.1 | 67.6 | 262 | 145 | 3 | <i>B. cinerea</i> glutamate oxaloacetate transaminase 2 (426 nt) |
| 242 | BC1G_07909 | 9.5 | 50.6 | 38.7 | 179 | 47 | 1 | <i>B. cinerea</i> hypothetical protein (447 nt) |
| 243 | BC1G_14663 | 10 | 34.3 | 30.5 | 63 | 62 | 2 | <i>B. cinerea</i> hypothetical protein (302 nt) |
| 244 | BC1G_05989 | 9.4 | 57.7 | 50.3 | 175 | 146 | 2 | <i>B. cinerea</i> ATP-citrate synthase subunit 1 (531 nt) |
| 245 | BC1G_15101 | 6.4 | 51.2 | 51.1 | 155 | 299 | 2 | <i>B. cinerea</i> hypothetical protein (470 nt) |
| 246 | BC1G_05989 | 9.4 | 57.7 | 50.3 | 180 | 94 | 1 | <i>B. cinerea</i> ATP-citrate synthase subunit 1 (531 nt) |

Table 1. Continued

| Spot no. | Acc. number | pI | MW | Cov (%) | Mascot MS | Mascot MS/MS | MS/MS peptides | Protein |
|----------|-------------|-----|------|---------|-----------|--------------|----------------|---|
| 247 | BC1G_14516 | 7.8 | 71.9 | 34.8 | 68 | 348 | 5 | <i>B. cinerea</i> hypothetical protein (633 nt) |
| 248 | BC1G_11336 | 6.4 | 89.3 | 52.2 | 304 | 125 | 2 | <i>B. cinerea</i> hypothetical protein (788 nt) |
| 249 | BC1G_15638 | 6.2 | 106 | 31.3 | 153 | 21 | 1 | <i>B. cinerea</i> translation elongation factor eEF-3 (947 nt) |
| 252 | BC1G_00471 | 4.8 | 55.4 | 50.8 | 108 | 73 | 2 | <i>B. cinerea</i> -conserved hypothetical protein (504 nt) |
| 252 | BC1G_00472 | 9.3 | 50.4 | 49.7 | 93 | x | x | <i>B. cinerea</i> -conserved hypothetical protein (465 nt) |
| 252 | BC1G_03668 | 6 | 68.7 | 39.6 | 88 | 131 | 2 | <i>B. cinerea</i> protein transport protein sec23s (621 nt) |
| 253 | BC1G_07152 | 5.4 | 48.6 | 55 | 194 | 152 | 3 | <i>B. cinerea</i> hypothetical protein (447 nt) |
| 254 | BC1G_07780 | 9.6 | 60.8 | 59.8 | 269 | 203 | 2 | <i>B. cinerea</i> hypothetical protein similar to mitochondrial ATP synthase alpha subunit (562 nt) |
| 255 | BC1G_15049 | 5.9 | 49.4 | 59.7 | 226 | 146 | 2 | <i>B. cinerea</i> saccharopine reductase (449 nt) |
| 257 | BC1G_05226 | 5.9 | 33 | 62.3 | 133 | 94 | 2 | <i>B. cinerea</i> hypothetical protein (300 nt) |
| 257 | BC1G_05617 | 5.7 | 50.4 | 58.5 | 172 | 169 | 2 | <i>B. cinerea</i> hypothetical protein (458 nt) |
| 258 | BC1G_09363 | 5.5 | 57.4 | 25.5 | 65 | 30 | 1 | <i>B. cinerea</i> hypothetical protein (514 nt) |
| 259 | BC1G_07012 | 5.9 | 49 | 65.3 | 116 | 36 | 1 | <i>B. cinerea</i> hypothetical protein (458 nt) |
| 259 | BC1G_07967 | 6.6 | 92.7 | 21.6 | 43 | 57 | 1 | <i>B. cinerea</i> hypothetical protein (850 nt) |
| 259 | BC1G_11228 | 8.9 | 61.2 | 71.2 | 282 | 181 | 2 | <i>B. cinerea</i> hypothetical protein (553 nt) |
| 260 | BC1G_02079 | 5.2 | 54.3 | 77.7 | 229 | 65 | 1 | <i>B. cinerea</i> hypothetical protein (503 nt) |
| 261 | BC1G_01271 | 6.4 | 33.1 | 66.3 | 181 | 142 | 2 | <i>B. cinerea</i> -conserved hypothetical protein (294 nt) |
| 262 | BC1G_01260 | 5.4 | 38.3 | 67.1 | 223 | 164 | 2 | <i>B. cinerea</i> -conserved hypothetical protein (359 nt) |
| 263 | BC1G_11347 | 5.6 | 62.8 | 56.7 | 202 | 113 | 1 | <i>B. cinerea</i> hypothetical protein (573 nt) |
| 266 | BC1G_07147 | 6.7 | 52.8 | 56.2 | 196 | 161 | 2 | <i>B. cinerea</i> hypothetical protein (475 nt) |
| 266 | BC1G_08475 | 5.4 | 67 | 41.6 | 112 | 178 | 2 | <i>B. cinerea</i> -conserved hypothetical protein (613 nt) |
| 267 | BC1G_14004 | 5.4 | 59.2 | 73.8 | 326 | 186 | 2 | <i>B. cinerea</i> hypothetical protein (545 nt) |
| 268 | BC1G_01594 | 6 | 39.3 | 31 | 63 | 114 | 2 | <i>B. cinerea</i> hypothetical protein similar to endochitinase (364 nt) |
| 269 | BC1G_01292 | 6.3 | 88.7 | 28.4 | 164 | 58 | 1 | <i>B. cinerea</i> hypothetical protein (805 nt) |
| 270 | BC1G_05448 | 5.7 | 31.8 | 45.7 | 181 | 130 | 2 | <i>B. cinerea</i> hypothetical protein (293 nt) |
| 271 | BC1G_03241 | 5.5 | 48.9 | 64.2 | 225 | 146 | 2 | <i>B. cinerea</i> -conserved hypothetical protein (450 nt) |
| 272 | BC1G_08581 | 5.3 | 43.3 | 56.7 | 161 | 151 | 2 | <i>B. cinerea</i> hypothetical protein (386 nt) |
| 272 | BC1G_13183 | 5.4 | 48 | 39.8 | 83 | 41 | 1 | <i>B. cinerea</i> hypothetical protein (450 nt) |
| 273 | BC1G_00939 | 5.3 | 47 | 59.4 | 232 | 215 | 2 | <i>B. cinerea</i> hypothetical protein (416 nt) |
| 275 | BC1G_10846 | 5.1 | 46 | 32.9 | 45 | 104 | 2 | <i>B. cinerea</i> hypothetical protein similar to hsp70 (425 nt) |
| 275 | BC1G_11821 | 9 | 61.6 | 50.5 | 168 | 266 | 2 | <i>B. cinerea</i> hypothetical protein (572 nt) |
| 275 | BC1G_14762 | 4.7 | 39.6 | 39.5 | 76 | 26 | 1 | <i>B. cinerea</i> D-ribulokinase (362 nt) |
| 276 | BC1G_02094 | 5.7 | 54.9 | 32.1 | 110 | 159 | 1 | <i>B. cinerea</i> hypothetical protein (489 nt) |
| 277 | BC1G_00027 | 6.2 | 29.8 | 56.7 | 141 | 248 | 2 | <i>B. cinerea</i> hypothetical protein (275 nt) |
| 278 | BC1G_00284 | 6.7 | 50 | 64 | 228 | 117 | 2 | <i>B. cinerea</i> hypothetical protein (456 nt) |
| 279 | BC1G_11963 | 5 | 24.8 | 56.6 | 141 | 136 | 2 | <i>B. cinerea</i> glycogen-branching enzyme (212 nt) |
| 280 | BC1G_12426 | 5.7 | 86.5 | 22.7 | 68 | 126 | 2 | <i>B. cinerea</i> predicted protein (774 nt) |
| 281 | BC1G_06095 | 5.8 | 51.1 | 57.6 | 222 | 143 | 2 | <i>B. cinerea</i> hypothetical protein (472 nt) |
| 282 | BC1G_05297 | 6.2 | 46.5 | 27.9 | 27 | 236 | 4 | <i>B. cinerea</i> hypothetical protein (408 nt) |
| 283 | BC1G_05327 | 6.1 | 133 | 47.6 | 383 | x | x | <i>B. cinerea</i> pyruvate carboxylase (1210 nt) |
| 284 | BC1G_11051 | 6.1 | 41.7 | 26.7 | 82 | x | X | <i>B. cinerea</i> hypothetical protein (401 nt) |
| 285 | BC1G_06849 | 4.8 | 43 | 19.8 | 104 | 169 | 1 | <i>B. cinerea</i> hypothetical protein similar to aspartic proteinase precursor (399 nt) |
| 286 | BC1G_02223 | 4.5 | 57.6 | 26.9 | 182 | 35 | 1 | <i>B. cinerea</i> hypothetical protein (532 nt) |
| 287 | BC1G_08294 | 4.7 | 30.9 | 35.1 | 96.5 | x | x | <i>B. cinerea</i> 14-3-3 protein homolog (276 nt) |
| 288 | BC1G_00383 | 6.3 | 24.6 | 41.6 | 68 | 104 | 1 | <i>B. cinerea</i> GTP-binding nuclear protein ran-1 (219 nt) |
| 289 | BC1G_04759 | 5.6 | 55 | 38.4 | 115 | 89 | 1 | <i>B. cinerea</i> -conserved hypothetical protein (511 nt) |
| 290 | BC1G_00121 | 5 | 45.1 | 31.3 | 104 | x | x | <i>B. cinerea</i> hypothetical protein (406 nt) |
| 291 | BC1G_13197 | 5.7 | 125 | 30 | 255 | x | x | <i>B. cinerea</i> L-aminoadipate-semialdehyde dehydrogenase large subunit (1131 nt) |
| 292 | BC1G_05327 | 6.1 | 133 | 43.3 | 432 | x | x | <i>B. cinerea</i> pyruvate carboxylase (1210 nt) |
| 293 | BC1G_08174 | 9.6 | 26 | 33.2 | 99 | x | x | <i>B. cinerea</i> hypothetical protein (235 nt) |
| 294 | BC1G_10819 | 5 | 30.4 | 26.3 | 90 | x | x | <i>B. cinerea</i> hypothetical protein (304 nt) |
| 295 | BC1G_02778 | 6.1 | 31.7 | 27.7 | 73 | x | x | <i>B. cinerea</i> hypothetical protein (285 nt) |
| 296 | BC1G_09983 | 4.6 | 23.2 | 19.7 | 44 | x | X | <i>B. cinerea</i> nascent polypeptide-associated complex alpha polypeptide (213 nt) |
| 297 | BC1G_11887 | 4.7 | 67.4 | 14.7 | 76 | x | x | <i>B. cinerea</i> hypothetical protein (605 nt) |

Table 1. Continued

| Spot no. | Acc. number | pI | MW | Cov (%) | Mascot MS | Mascot MS/MS | MS/MS peptides | Protein |
|----------|-------------|-----|------|---------|-----------|--------------|----------------|--|
| 298 | BC1G_03007 | 6.2 | 40.3 | 36.6 | 100 | x | x | <i>B. cinerea</i> hypothetical protein (358 nt) |
| 299 | BC1G_15149 | 5.3 | 109 | 16.1 | 112 | x | x | <i>B. cinerea</i> cytoskeleton assembly control protein (962 nt) |
| 300 | BC1G_05074 | 7.8 | 34.5 | 29.7 | 124 | x | x | <i>B. cinerea</i> hypothetical protein similar to succinyl-CoA synthetase (330 nt) |
| 301 | BC1G_05327 | 6.1 | 133 | 43.4 | 295 | x | x | <i>B. cinerea</i> pyruvate carboxylase (1210 nt) |
| 302 | BC1G_06400 | 6.4 | 47.7 | 57.7 | 173 | x | x | <i>B. cinerea</i> -conserved hypothetical protein (440 nt) |
| 303 | BC1G_07147 | 6.7 | 52.8 | 16.2 | 64.6 | x | x | <i>B. cinerea</i> hypothetical protein (475 nt) |
| 304 | BC1G_01161 | 6.2 | 53 | 18.4 | 70 | x | x | <i>B. cinerea</i> hypothetical protein (499 nt) |
| 305 | BC1G_00558 | 5.9 | 16 | 34.8 | 85 | x | x | <i>B. cinerea</i> superoxide dismutase Cu-Zn (155 nt) |
| 306 | BC1G_06628 | 6.5 | 67.2 | 12.9 | 54 | x | x | <i>B. cinerea</i> hypothetical protein similar to septin 2 (588 nt) |

Acc Number, accession number of the identified protein in the *B. cinerea* database from the Broad Institute (www.broad.mit.edu); pI, isoelectric point (theoretical); MW, molecular weight (theoretical); Cov, percent of the residues in each protein sequence that has been identified (from PMF when a PMF identification was successful); Mascot MS, Mascot score of PMF identification; Mascot MS/MS, Mascot score of the peptide fragmentation fingerprint identification; MS/MS peptides, number of peptides from MS/MS search.

buffered phenol, pH 8.0 (Biomol, Hamburg, Germany) was added, and the resulting mixture was vortexed for 30 s. The phenol phase was separated by centrifugation and transferred to a fresh tube. After addition of five volumes of cold 0.1 M ammonium acetate in methanol, the proteins were precipitated from the phenol phase overnight at -20°C . The precipitated proteins were recovered by centrifugation, washed twice with cold 0.1 M ammonium acetate in methanol and twice with 80% v/v acetone. The final pellet was air-dried and stored at -80°C . A separately precipitated aliquot was dissolved in a small volume of 8 M urea for protein quantification using the Bradford assay (Bio-Rad, München, Germany) with BSA as standard. The few 2-DE proteomic approaches carried on *B. cinerea* mycelia proteins were made on the basis of an extraction protocol based on phosphate buffer solubilization and TCA/acetone precipitation [3, 5]. However, the combined use of TCA precipitation and phenol extraction improved better spot definition, reduced streaking and led to a higher number of detected spots (data not shown). 2-DE was performed as follows: the first dimension was run using 17 cm IPG strips (3–10 non-linear pH gradient, Amersham) on a Bio-rad Protean IEF cell according to the manufacturer's instructions. The second dimension was run on a Protean II xi Cell (Bio-rad) using self-cast 14% acrylamide SDS-PAGE gels [14]. The gels were stained with Coomassie PAGE Blue (Fermentas, St. Leon-Rot, Germany) or Sypro Ruby (Invitrogen, Paisley, UK) stains according to the manufacturer's instructions (Fig. 1). Only those spots that were present in all the replicates were selected for identification process.

2-DE gels of mycelia cultured with glucose displayed a characteristic profile with 262 ± 18 detected spots, whereas gels of mycelia cultured with cellulose showed 318 ± 5 (data not shown), indicating that the use of a particular carbon source, such as glucose or cellulose, induces a characteristic fungal response. These differences will be analyzed in a further study (manuscript in preparation).

Protein spots were selected visually and robotically excised from Coomassie-stained gels and digested using the Proteiner spII and dp systems (Bruker Daltonic, Bremen,

Germany). Following robotic target preparation, PMF data were collected on an UltraflexIII MALDI ToF/ToF spectrometer (Bruker). LIFT MS/MS spectra [15] were then collected on selected precursors for verification of PMF identifications and further elucidation. Both MS and MS/MS data were used to search against the *B. cinerea* protein database (<http://www.broad.mit.edu>/ August 2007 release) or the NCBI non-redundant database (<http://www.ncbi.nlm.nih.gov> May 2007 release) using MASCOT (<http://www.matrixscience.com>). MS acquisition and database search parameters are available in Supporting Information. Further experimental details are available from the authors upon request.

In constructing the 2-DE map of *B. cinerea*, 267 spots excised from the gel yielded protein identifications by MALDI-TOF/TOF MS/MS analysis. The identifications represent 303 positive hits to proteins and their respective isoforms (Table 1), representing approximately 2% of the predicted proteins in the annotated genome. This information is available in the public database World-2DPAGE (www.expasy.org/world-2dpag/) for the *B. cinerea* research community (Accession number: 0005-“*B. cinerea* mycelium cultured in carboxymethyl cellulose”). Proteins with an important role during plant infection have been identified (Table 1) (*i.e.* spot 14 – *B. cinerea* peptidyl-prolyl *cis*–*trans* isomerase) [16]. Moreover, other proteins with a suggested role in the infection process are listed too (*i.e.* spot 156 – glyceraldehyde 3-phosphate dehydrogenase) [3, 17]. However, a large number of the identified proteins were annotated as “*B. cinerea* hypothetical proteins.” These proteins were annotated by BLAST searches, and the results are listed in Supporting Information Table 2.

Identified proteins from *B. cinerea* mycelia were distributed into categories according to their molecular functions and involvement in biological processes based on PANTHER classification system (www.pantherdb.org/). Classification by molecular function showed that more than 40% of the proteins fell into three categories: (i) oxidoreductase 20%, (ii) nucleic acid binding 12% and (iii) synthase and synthetase 10% (Fig. 2A). Oxidoreductase activity (catalysis of redox reactions) has been

well documented in *B. cinerea*. During infection process, AOS seem to play an important role. Plants produce a lot of AOS as a defense mechanism, but the fungus seems to produce these radicals as well. However, the fungus also produces free radical scavenger proteins and metabolites that protect it from AOS attack, such as oxalic acid, laccase, catalase and several peroxidases [18]. Moreover, some of these proteins have been directly related to virulence and toxin production [3, 19]. Nucleic acid-binding activity implies nucleotide-binding enzymes or DNA-binding proteins. Among other functions, the relevance of such proteins in the cascade of signals has been deeply studied in

B. cinerea [20]. Ten percent of identified proteins (synthase and synthetase) have a role in the synthesis of several molecules, especially nucleoside triphosphates. Other significantly represented categories include transferases (8%), hydrolases, lyases and ligases (6%). Most of the proteins (45%) categorized by biological process (Fig. 2B) were grouped into three categories: (i) protein metabolism and modification (21%), (ii) carbohydrate metabolism (15%) and (iii) amino acid metabolism (9%). One of the goals of proteomic approaches is to study the processes related with protein modifications. Metabolic processes that involve protein synthesis, modification or degradation may

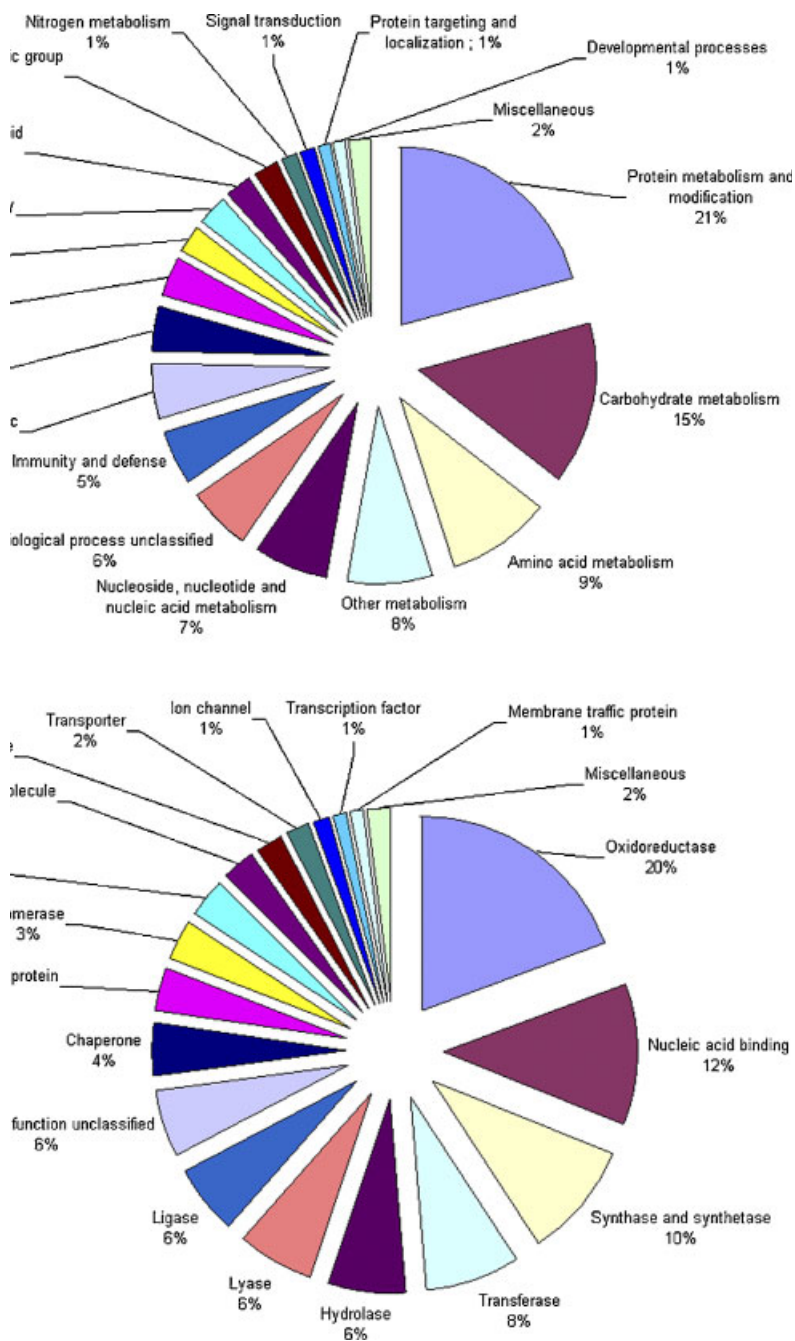


Figure 2. Representations of the distribution of identified *B. cinerea* proteins according to their (A) molecular function and (B) biological process. Categorizations were based on information provided by the online resource PANTHER classification system.

confer multiple biological functions to individual genes [21]. Thirty percent of the identified proteins are related with the biosynthesis, assembly, folding, translational regulation of proteins or amino acid biosynthesis transport, *etc.* The role of these modifications and their impact on virulence, intracellular signal cascades and other biological processes is an area for further investigation. Proteins related to carbohydrate metabolism (15%) are involved in the use of CMC as a sole carbon source. Since CMC is one of the major components of the plant cell wall, many of them may have a crucial role in the pathogenicity process.

In summary, this study describes the first proteomic analysis of the phytopathogenic fungus *B. cinerea*, containing an important number of identified proteins. Moreover, the use of CMC as a sole carbon source may reveal proteins involved in the infection cycle such as pathogenicity or virulence factors. The identification of these proteins, their PTMs, isoforms and subunits will provide a new source of knowledge to unravel the molecular basis of pathogenesis. Moreover, this *B. cinerea* 2-DE proteomic map can be a hypothesis driver for fungicide design based on new therapeutical targets. At present, there is no fungicide on the market that was developed on the basis of targeted molecular research [4]. Some of the identified proteins may also be used to develop a procedure for field diagnosis of plant infections [22].

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References

- [1] Elad, Y., Williamson, B., Tudzynski, P., Delen, N., *Botrytis: Biology, Pathology and Control*, Kluwer Academic Publishers. Dordrecht, The Netherlands 2004.
- [2] Fernández-Acero, F. J., Carbú, M., Garrido, C., Vallejo, I., Cantoral, J. M., Proteomic advances in phytopathogenic fungi. *Curr. Proteomics* 2007, 4, 79–88.
- [3] Fernández-Acero, F. J., Jorge, I., Calvo, E., Vallejo, I. *et al.*, Proteomic analysis of phytopathogenic fungus *Botrytis cinerea* as a potential tool for identifying pathogenicity factors, therapeutic targets and for basic research. *Arch. Microbiol.* 2007, 187, 207–215.
- [4] Williamson, B., Tudzynski, B., Tudzynski, P., Van Kan, J. A. L., *Botrytis cinerea*: the cause of grey mould disease. *Mol. Plant Pathol.* 2007, 8, 561–580.
- [5] Fernández-Acero, F. J., Jorge, I., Calvo, E., Vallejo, I. *et al.*, Two-dimensional electrophoresis protein profile of the phytopathogenic fungus *Botrytis cinerea*. *Proteomics* 2006, 6, S88–S96.
- [6] Myung, K., Hamilton-Kemp, T. R., Archbold, D. D., Interaction with and effects on the profile of proteins of *Botrytis cinerea* by C6 aldehydes. *J. Agric. Food Chem.* 2007, 55, 2182–2188.
- [7] Van Kan, J. A. L., Licensed to kill: the lifestyle of a necrotrophic plant pathogen. *Trends Plant Sci.* 2006, 11, 247–253.
- [8] Choquer, M., Fournier, E., Kunz, C., Levis, C. *et al.*, *Botrytis cinerea* virulence factors: new insights into a necrotrophic and polyphageous pathogen. *FEMS Microbiol. Lett.* 2007, 277, 1–10.
- [9] Tournu, H., Serneels, J., Van Dijck, P., Fungal pathogens research: novel and improved molecular approaches for the discovery of antifungal drug targets. *Curr. Drug Targets* 2005, 6, 909–922.
- [10] Dhingra, V., Gupta, M., Andacht, T., Fu, Z. F., New frontiers in proteomics research: a perspective. *Int. J. Pharm.* 2005, 299, 1–18.
- [11] Vallejo, I., Carbú, M., Muñoz, F., Rebordinos, L., Cantoral, J. M., Inheritance of chromosome-length polymorphisms in the phytopathogenic ascomycete *Botryotinia fuckeliana* (anam. *Botrytis cinerea*). *Mycol. Res.* 2002, 106, 1075–1085.
- [12] Yajima, W., Kav, N. N. V., The proteome of the phytopathogenic fungus *Sclerotinia sclerotiorum*. *Proteomics* 2006, 6, 5995–6007.
- [13] Röhrig, H., Schmidt, J., Colby, T., Bräutigam, A. *et al.*, Desiccation of the resurrection plant *Craterostigma plantagineum* induces dynamic changes in protein phosphorylation. *Plant Cell Environ.* 2006, 29, 1606–1617.
- [14] Laemmli, U. K., Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature* 1970, 227, 680–685.
- [15] Suckau, D., Resemann, A., Schuerenberg, M., Hufnagel, P. *et al.*, A novel MALDI LIFT-TOF/TOF mass spectrometer for proteomics. *Anal. Bioanal. Chem.* 2003, 376, 952–965.
- [16] Viaud, M. C., Balhadere, P. V., Talbot, N. J., A *Magnaporthe grisea* cyclophilin acts as a virulence determinant during plant infection. *Plant Cell* 2002, 14, 917–930.
- [17] Alderete, J. F., Millsap, K. W., Lehker, M. W., Benchimol, M., Enzymes on microbial pathogens and *Trichomonas vaginalis*: molecular mimicry and functional diversity. *Cell. Microbiol.* 2001, 3, 359–370.
- [18] Gil-ad, N. L., Bar-Nun, N., Noy, T., Mayer, A. M., Enzymes of *Botrytis cinerea* capable of breaking down hydrogen peroxide. *FEMS Microbiol. Lett.* 2000, 190, 121–126.
- [19] Siewers, V., Viaud, M., Jimenez-Teja, D., Collado, I. G. *et al.*, Functional analysis of the cytochrome P450 monooxygenase gene *bcbot1* of *Botrytis cinerea* indicates that botrydial is a strain-specific virulence factor. *Mol. Plant Microbe Interact.* 2005, 18, 602–612.
- [20] Tudzynski, B., Gronover, C. S., in: Elad, Y., Tudzynski, B. W. P., Delen, N. (Eds.), *Botrytis: Biology, Pathology and Control*, Kluwer Academic Publishers, Dordrecht 2004, pp. 85–97.
- [21] Graves, P. R., Haystead, T. A. J., Molecular biologist's guide to proteomics. *Microbiol. Mol. Biol. Rev.* 2002, 66, 39–63.
- [22] Padliya, N. D., Cooper, B., Mass spectrometry-based proteomics for the detection of plant pathogens. *Proteomics* 2006, 6, 4069–4075.